



Hepatotoxicity, Hyperglycemic and hyperlipidemic effect of Aqueous Leaf Extract of *Nicotianatabaccum* in male Wistar albino rats

Nweze Chibuzo Carole¹, Nweze Otitoadaeze²

The leaves of *Nicotianatabaccum* is used as a medicinal plant in Northern Nigeria. The present study aimed at evaluating the biochemical effects of aqueous leaves extracts of *Nicotianatabaccum* in liver indices, blood glucose and some lipid profile in male Wistar albino rats. The animals were divided into four groups: Group 1 was treated with normal saline, group 2, 3 and 4 with 0.5mL of the extracts, at 150 mg/kg, 300 mg/kg and 450 mg/kg respectively. The treatment was given daily for fourteen days. Blood samples were collected and assayed. The results showed significantly ($P<0.05$) elevated blood glucose level accompanied by significant increase in alanine amino transferase (ALT), aspartate aminotransferase (AST) and alkaline phosphatase (ALP) activities in the treated groups (1,2, and 3) when compared with group 1. Bilirubin (total and direct) showed a significant ($P<0.05$) increase in concentration in groups 2, 3, and 4 with the total bilirubin having the highest increase in the activity compared to group 1. Total cholesterol, triglyceride and low density lipoproteins were significantly elevated in groups 2, 3, and 4; while high density lipoprotein was significantly ($p<0.05$) reduced compared to group 1. The leaves extract at different concentrations (doses) showed that group 4 (600mg/kg) had a significant ($p<0.05$) increase in blood biochemical samples compared to lower concentrations in group 2 and 3. Also group 3 showed a significant ($p<0.05$) increase compared to group 2. Therefore, the present studyrevealed that the aqueous leaves extracts of *Nicotianatabaccum* might not be safe for human consumptions as they are associated with liver toxicity, hyperglycemia and hyperlipidemia.

INTRODUCTION

In recent years, medicinal plants occupy an important position for being the paramount sources of the discovery of pharmacologically active compounds (Mustapha, 2014). Many medicinal plants use traditionally have been in use by man without the actual knowledge of their toxic potential. In Nasarawa and other parts of northern Nigeria, one such plants used is tobacco plant leaves (*Nicotianatabaccum*), are used in two major forms which is the smoked and the smokeless, the tobacco snuff and chewing tobacco (Aduema et al., 2012), used as an organic pesticide and in the form nicotine tartrate in medicine (Ugbor et al., 2013). Tobacco snuff is the powdered form blended with potash as the main additive in Nigeria (Ureme et al., 2007) and has been recommended as a substitution for nicotine in cigarette since it is devoid of hazardous elements such as tar and carbon monoxide (Russel et al, 1980). Large number of people believes that using smokeless tobacco is safer than smoking it (Ugbor et al., 2013). This however, is not true because smokeless tobacco can induce addiction to nicotine and leukoplakia (Armitage et al., 1978; Armitage and Turner, 1970).

Nicotianatabaccum (Family Solanaceae), is a small perennial herbaceous plant with long leaves, flowers and seeds that is the most

commonly grown of all plants in the Nicotiana genus. It is widely grown in Northern Nigeria and Sudan. The leaves of the plants are commercially used and processed into tobacco products (Ren and Timko, 2001).

Tobacco has many constituents, but nicotine is singled out as having the highest concentration and most immediate pharmacological action (Heishman et al., 1994). Nicotine is extremely toxic, about as toxic cyanide, and only 60 mg are needed to kill humans but while smoking tobacco, it contain a small portion of nicotine which the body metabolize to a non-toxic substance (Dasofunjo et al., 2014). Nicotine has major central nervous system (CNS) stimulant, though these effects are not as intense as what is observed with cocaine and amphetamines but nicotine enhance effect on alertness, learning, and memory (Heishman et al., 1994). Unlike many other plants Solanaceae family, do not contain tropane alkaloids, which are reported to be poisonous to humans and other animals (Fafioye and John-Dewole, 2013). However, other compounds such as germacrene, anabasine and other piperidine alkaloids (varying between species), which are powerful enough to deter most herbivores (Benowitz, 1990). A number of such animals have developed the ability to feed on Nicotiana species without being harmed (Fafioye and John-Dewole, 2013). Nonetheless, tobacco is unpalatable to many species, and therefore some tobacco plants such as *Nicotianaglauca* have become established as invasive weeds in some species (Fafioye and John-Dewole, 2013). Tobacco either smoked or

¹Department of Biochemistry and Molecular Biology, Nasarawa State University, Keffi, Nigeria; Email: chibuzoihe@gmail.com; ²Department of Medicine and Surgery, Anambra State University Teaching Hospital Awka, Nigeria; Email: nwezeotito@gmail.com

smokeless contains nicotine and other metabolites such as tabacine, tabacine, 4-(methylnitrosamino)-1-(3-pyridyl)-1-butanone), 2-Naphylamine, 4-methyl-nitrosamino)-4-(3-pyridyl)-butanal (NNA), 2,3,6-Trimethyl-1,4-naphthoquinone, 2-Methylquinone, N-nitrosornicotine, Cadmium (Cd), mercury (Hg), Propionic acid, Anatabine, Nicotelline, Choline, Pyrene, Cembrene and Choline which has been implicated with tobacco associated cancers and diseases (Adedayo et al., 2011; Stepanov et al., 2010; Addo et al., 2008; Hecht et al., 2007; Cerami et al., 1997; Chiba and Masironi, 1992; Hecht and Hoffmann, 1988; Hoffmann and Hecht, 1985; Hecht et al., 1978). These effects may account for part of nicotine reinforcing effects in humans. Nicotine affects cardiovascular system through its an autonomic effect (Checkoway et al., 2002).

The stimulation of the heart and its resultant increased demand for oxygen underlie the association of nicotine and disease resulted. Moreover, when there is a failure to deliver an adequate supply of oxygen to the heart, it may result in chest pain (angina) or a heart attack (Carlisle et al., 2004) due to stimulation of sympathetic ganglia and adrenal medulla combined with discharge of catecholamines (Fafioye and John-Dewole, 2013). One of the famous acute effects of nicotine is its relationship to lower body weight (Julien, 1996). In this regard, nicotine decrease appetite for sweet foods and increase the amount of energy the body uses both while resting and exercising (Vik et al., 1996). These features of nicotine helps to explain the research which show that smokers tend to weigh less compare to non-smokers. On the other hand, quitting from smoking is associated with weight gain (Meyer et al., 1997). Tobacco has a long history of use by traditional medical practitioners as a relaxant, but because of its highly addictive property, it is seldom employed internally to treat depression and heart diseases or externally to treat rheumatic swelling, skin diseases and scorpion stings (Fafioye and John-Dewole, 2013).

However, every drug is associated with hepatotoxicity almost certainly due to its ability to generate free radicals and to cause disturbance in hepatocyte biochemistry (Fernandez-checa and Kaplowitz, 2005). However, the increasing demand and consumption of Nicotine, either smoked or chewed has highlighted researchers to assess its toxicological effect in the liver since the liver is responsible for the detoxification of foreign substances. Diabetes mellitus refers to a group of metabolic diseases characterized by high blood glucose (hyperglycaemia) concentration result from defects in insulin secretion, or action. It is a morbid condition with high prevalence worldwide (Etuk, 2010). Hyperlipidemia has been shown to be associated with coronary heart diseases and oftentimes are associated with diabetes mellitus (Lotfy *et al.*, 2013).

MATERIALS AND METHODS

Plant Material

Fresh tobacco plant leaves were collected from Nasarawa Eggon of Nasarawa State which was identified at the herbarium of the Plant Science and Biotechnology Unit, Department of Biological Sciences of Nasarawa State University, Keffi.

Preparation of Extract

Fresh plant leaves were allowed to dry at room temperature which was letter grinded to powder form using mortar and pestle. 100 g of the powdered extract were soaked into 1000ml of distilled water for 24 h (Mustapha et al., 2014). After 24 h, the soaked extract was filtered with sieve and then Whatman filter paper (No 1) was used to re-filter it and pure extract was obtained. The residue was discarded. The filtered

extract was subjected to water bath for evaporation at 98°C. Solid extract was obtained which was used for analysis.

Experimental Design

Male albino rats weighing (170-175g) were purchased from the animal house of the National Veterinary Research Institute, Vom, Jos, Plateau State. The animals were acclimatised to environmental condition for a week prior to the experiment and had access to food (rat chow) and distilled water.

The animals were grouped into four groups of five rats each. The first group were orally administered normal saline and were sacrifice 24 h after the last administration. This group represents the control animals. The second groups were given 0.5mL of aqueous leaves extract dose of 150 mg/kg body weight was orally administered and the animals were sacrifice 24 h after the last administration. The third groups were administered aqueous leaves extract dose of 300 mg/kg body weight was orally administered and the animals were sacrifice 24 h after the last administration. The fourth group were orally administered with an aqueous leaves extract dose of 450 mg/kg body weight was orally administered and the animals were sacrifice 24 h after the last administration. However, the treatment periods lasted for fourteen days.

The blood was collected as previously described by the Institutional Animal Care and Use Committee (IACUC). Retro-orbital blood collection method was employed. Briefly, the animals were fully anesthetised by a drop of proparicaine in each eye to minimise discomfort. The animal was held by the back of the neck and the loose skin of the head was tightened with thumb and middle finger to keep the animal stable. The tip of non-heparinised capillary tube was place at the medial can thus of the eye under the nictitating membrane. With gentle thrust and rotation motion past the eyeball the tube will enter the slightly resistant sinus membrane and the eye ball remained uninjured. As soon as the sinus was punctured, the blood enters the tubing was capillary action. The blood was allowed to clot and then centrifuged at 3500 rpm for 5 min using table centrifuge. The serum was then separated from the blood and further used for analysis.

Biochemical Analysis

The serum separated was used to assay for liver enzymes; alanine aminotransferase (ALT) and aspartate aminotransferase (AST) were assayed by the method described by Reitman and Frankel, (1957), alkaline phosphatase (ALP) was also analysed by method of Rec (1972). Serum Bilirubin was determined using colorimetric method as previously described by Jendrassik and Grof, (1938). The serum glucose and total cholesterol (TC) were determined by the method of Trinda, (1969); Searcy and Berquist, (1960) respectively. High density lipoprotein cholesterol (HDLc) and triglyceride (TG) were determined by the method of Friedwald *et al.* (1972); Assman *et al.* (1984) respectively. Low density lipoprotein cholesterol (LDLc) was calculated using formula (Albers *et al.*, 1978)

$$LDL = TC - TG/5 - HDL$$

Statistical Analysis

Statistical analysis of the result analysed in one-way analysis of variance ANOVA using Smith's Statistical Package, followed by Student t-test for comparison. Statistical significant difference was set at $p < 0.05$.

RESULTS AND DISCUSSION

The measurement of activities of enzymes in tissues and body fluids plays a significant role in the investigation and diagnosis of diseases

Table 1 Effect of aqueous leaves extract of *Nicotianatabacum* on Some selected serum liver indices in male Wister rats

Treatment	ALT (μ/L)	AST (μ/L)	ALP (μ/L)	TB (μmol/L)	DB (μmol/L)
(Mgkg ⁻¹)					
Control	12.4±4.32	11.6 ± 5.37	137.3 ± 9.54	5.7 ± 2.64	17.9 ± 5.16
150 (AE)	25.5±3.71*	28.6 ± 7.20	*289.9 ± 13.07*	10.5 ± 4.43*	28.3 ± 7.00*
300 (AE)	49.8±2.96 ^a	49.00 ± 4.81*	^a 342.0 ± 6.75 ^a	11.6 ± 3.50	39.6 ± 7.45 ^a
450 (AE)	66.4±5.34 ^b	70.3 ± 6.62*	^b 394.6 ± 1.43*	^b 26.7 ± 6.28*	^b 51.1 ± 4.60 ^b

Data expressed as mean ± SD, *Significant different from the control (P<0.05), (n=5); ^a significantly different from group 1 150(AE); ^b significantly different from group 2 300 (AE)

Table 2 Effect of aqueous leaves extract of *Nicotianatabacum* on serum glucose of male Wister rats

Treatment	Serum glucose (mg/Dl)	Serum glucose (mg/Dl)
	at 7 days	at 14 days
Control	72.62±22.61	75.03±43.51
150 (AE)	98.93±10.35*	121.85±50.06 ^c
300 (AE)	121.06±18.27 ^a	163.29±17.02 ^{ac}
450 (AE)	140.22±19.03 ^b	201.68±64.90 ^{bc}

Data expressed as mean ± SD; n – 5; *Significant different from the control (P<0.05), (n=5); ^a significantly different from group 1 150(AE); ^b significantly different from group 2 300 (AE); ^c different concentrations significantly different from serum glucose level at 7days

Table 3 Effect of aqueous leaves extract of *Nicotianatabacum* on serum lipid profiles (mmol/L) of male Wister rats

Treatment (Mgkg ⁻¹)	Cholesterol	Triglyceride	HDLcLDLc	
Control	2.98±0.05	2.36±0.02	1.98±0.02	0.69±0.01
150 (AE)	4.64±0.02*	3.71±0.05*	0.92±0.05*	1.16±0.03*
300 (AE)	5.86±0.07 ^a	4.39±0.06 ^a	1.08±0.07*	1.44±0.05 ^a
450 (AE)	6.00±0.03 ^b	4.98±0.09 ^b	0.71±0.04 ^b	1.68±0.08 ^b

Data expressed as mean ± SD, *Significant different from the control (P<0.05), (n=5); ^a significantly different from group 1 150(AE); ^b significantly different from group 2 300 (AE)

(Molowo, 2000). The liver plays a key role in many metabolic processes. This fact demonstrates the biochemical importance of the organ, severe hepatic injury as a result of the metabolism of some toxic phytochemicals found in plants and failure of metabolic produces to be found in plants and the metabolic products to be eliminated by the liver (Giedan et al., 2004) may be associated with distortion of these function thereby leading to increase in the serum concentration of Aspartate aminotransferase (ASP), Alanine aminotransferase (ALT) and alkaline phosphatase activities (ALP) as found in the present study.

ALP is widely distributed within different tissues possessing one or more of the isoenzymes in bone, liver and intestine (Whitby et al., 1989). ALP is useful in diagnosis hepatobiliary lesions and Osteoblastic bone diseases (Wolf, 1978). Elevated activities of liver ALP immediately after the administration of the extract implied activation of the enzyme molecule. However, since there was significant change in the levels of serum ALP, the elevation in the liver ALP activity could be therefore attributed to enzyme activation, ALT, is a more specific enzyme of liver damage than AST. This is probably as a

result of induction of enzyme synthesis due to defence reaction of the organ to the exogenous substances or loss of other proteins (Akanji, 1989). This trend may have consequential effect on amino acid metabolism.

The result in Table 1 showed that there is significant increase in alkaline phosphatase in all treated groups compared to the control. Elevated levels indicate that there was an impaired bile formation (Cholestasis), intrahepatic cholestasis or infiltrative disease of the liver.

The ALT and AST are useful indicators for identifying inflammation and necrosis of the liver (Tikian et al., 1979). ALT has the highest concentration in the liver while AST has a lesser activity concentration in the kidney and skeletal muscle (Tikian et al., 1979). The activity of AST is located in the microsomal and mitochondrial portions of the liver cells as well as in the skeletal and cardiac muscles, pancreas and kidney (Whitby et al., 1989). ALT measurements are more liver specific than the AST and its activity is usually greater at early or acute hepatocellular diseases (Whitby et al., 1989). A marked elevation of ALT, however in the presence of mild to moderate elevation of AST

indicate either hepatic disease or hepatic disease combined with other conditions (Tikian et al., 1979). The result of this study showed significant increase in the activity of both ALT and AST as compared to the control in all the groups treated (Table 1). Thus, this finding did not provide evidence of clinical safety of the plant at the higher concentrations as indicated by the results. The significant increase of serum total and direct bilirubin, indicate defective liver excretory function (Chessbrough, 1991) and impaired synthetic function of the liver (Harold et al., 1980). The result of these findings in liver enzymes is in accordance with the one reported elsewhere (Mitra et al., 1980) on seed aqueous extract of *Nicotianatobaccum*.

Diabetes mellitus is a metabolic disease characterized by high blood glucose (hyperlipidemia) level. In the present study, administration of aqueous extract of *Nicotianatobaccum* in different concentrations of 0.5mL in 150mg/kg, 300mg/kg and 450mg/kg body weight (Table 2) to the rats caused significant ($p < 0.05$) elevation of serum blood glucose level as compared to control. This can be as result of defects in the insulin secretion. *Nicotianatobaccum* is a Beta-cytotoxic agents and it caused beta-cytotoxic action that is associated with the sudden release of insulin leading to severe transient hypoglycaemia followed by chronic hyperglycaemia resulting from the complete destruction of the pancreatic β -cells that secrete insulin. The finding in this study is in agreement with other studies that an agent of beta-toxicity impairs the release of insulin (Nweze et al., 2017).

The serum lipids level is usually raised in diabetes and present a risk factor for the development of cardiovascular diseases most especially coronary heart diseases (Luka et al., 2013). Administration of aqueous extract of *Nicotianatobaccum* in different concentrations of 0.5mL in 150mg/kg, 300mg/kg and 450mg/kg body weight (Table 3) showed significant ($p < 0.05$) increase in cholesterol (CHOL), triglyceride (TG) and low density lipoprotein (LDLc), with significant ($p < 0.05$) reduction in high density lipoprotein (HDLc). Diabetic states affect insulin and thus, may lead to inadequate utilization and activations of hormone-sensitive lipases, glucagon and catecholamines which in turns increase the breakdown of lipids and mobilization of free fatty acids from the peripheral depots (Yakubu et al, 2003). Activation of 3-hydroxy-3-methylglutaryl coenzyme responsible for the biosynthesis of cholesterol due to insulin deficiency might have been responsible for the observed hyperlipidemia. The hyperlipidemic effect of was compared to other studies that bark of the stem (Mustapha et al., 2014)

Furthermore this study has found that effect of nicotine in the serum is dose dependent. However these factors need further justification by further studies. For the reason that the method administration used by this study is oral, other ways of administration might be more toxic to the liver and other cells. Therefore to determine the toxicological effect of this study at moderate level, it oral administration method was chosen. Finally it was to compare its effect with other part of the world, thus the results were found to be comparable with that in the rest of the world as explained in the discussion section. It may be apparent to suggest that the plant leaves extract may not be safe for humans especially at higher concentrations.

REFERENCES

- Addo MA, Gbadago JK, Affum HA, Adom T, Ahmed K, Okley GM. 2008. Mineral profile of Ghanaian dried tobacco leaves and local snuff: A comparative study. *Journal of Radioanalytical and Nuclear Chemistry*, 277(3): 517-524.
- Adedayo AD, Tijani, AA, Musa AA, Adeniyi, T.D. (2011): Histological study of smoke extracts of *Tobacconicotiana* on the heart, liver, lungs, kidney, and testes of male Sprague-Dawley rats. *Nigeria Medical Journal*, 52(4): 217-222.
- Aduema W, Lelei SA, Osim EE, Koikoibo W, Nneli RO. 2012. Effect of chronic consumption of powdered tobacco (snuff) on anxiety, fear and social behaviours. *International Journal of Basic Applied and Innovative Research*, 1(4): 161-169.
- Akanji MA. 1986. A comparative biochemical study of the interaction of some typanosided with rat tissues cellular system. PhD Thesis, University of Ife, Ile-Ife.
- Albers JJ, Warmick GR, Cheung MC (1978) Quantitation of high density lipoprotein lipids, 13: 926-932
- Armitage A, Dollery C, Houseman T, Kohner E, Lewis PJ, Turner D. 1978. Absorption of nicotine from small cigars. *Clinical Pharmacology and Therapeutics*, 23: 143-151.
- Armitage AK, Turner DM. 1970. Absorption of nicotine in cigarette and cigar smoke through the oral mucosa. *Nature*, 226: 1231-1232.
- Assman G, Jabs HU, Kohnert U, Nolte W, Schriewer H. (1984) *Clinical Chemistry Acta*, 140: 77-87
- Carlisle DC, Hopkins TM, Gaither-Davis A, Silhanek MJ, Luketich JD, Christie NA, Siegfried JM. 2004. Nicotine signals through muscle-types and neuronal nicotinic acetylcholine receptor in both human bronchial epithelial cells and air way fibroblast. *Respiratory Research*, 5: 27.
- CeramiC, Founds H, Nicholl I, Mitsuhashi T, Vanpatten S. 1997. Tobacco Smoking is Source of Toxic Reactive Glycation Products. *Proceedings of the National Academy of Sciences of the United States of America*, 94(25): 15-20.
- Checkoway H, Powers K, Smith-Weller T, Franklin GM, Longstreth WT Jr, Swanson PD. 2002. Parkinson's disease risk associate with cigarette smoking. Alcohol consumption, and caffeine intake. *American Journal of Epidemiology*, 155: 732-738.
- Cheesbrough M. 1991. *Medical Laboratory Manual for Tropical Countries*. 2nd edn. Pp. 508-511, Cambridge, ELBS.
- Dasofunjo K, Okwari OO, Obembe AO, Olatunji TL, Ezugwu HC, Osim EE. 2014. Assessment of the Effect of *Cannabis sativa* and *Nicotianatobaccum* Leaves on some Haematological and Liver Function Indices of Albino Rats. *Journal of Pharmacy and Biological Sciences*, 9(1): 33-40.
- Fafioye O, John-Dewole, O. 2013. Toxicity of the aqueous and alcoholic Extracts of *Nicotiana Tabacum* (Tobacco Plant) on Histopathological and Haematological Parameters of Albino Rats. *Academia Arena*, 5(8): 24-29.
- Fernandez-Checa JC, Kaplowitz N. 2005. Hepatic mitochondrial glutathione: Transport and role in disease and toxicity. *Toxicology and Applied Pharmacology*, 204(3): 263-273.
- Friedwald WT, Levy RT, Fredrickson DS (1972) Estimation of the concentration of low density lipoprotein cholesterol in plasma without use of preparative ultra-centrifuge *clinchem* 18: 499-502.
- Giedan M, Pakman A, Laminu H. 2004. Effects of aqueous stem bark extract of *Monordicabalsaminlinin* on serum electrolytes and some haematological parameters in Normal and alcohol fed rats. *Pakistan Journal of Biological Sciences*, 7(8): 140-143.
- Harold V, Alan HG, Manurice B. 1980. *Practical Clinical Biochemistry*. pp. 890-913, London, William Heinemann.
- Hecht SS, Chen CB, Orna RM, Jacobs E, Adams JD, Hoffman D. 1978. Reaction of Nicotine and sodium nitrate: Formation of Nitrosamines and fragmentation of the pyrrolidine ring. *The Journal of Organic Chemistry*, 43:72.
- Hecht SS, Hoffmann D. 1988. Tobacco-specific nitrosamines, an important group of carcinogens in tobacco and tobacco smoke. *Carcinogenesis*, 9(6): 875-884.
- Heishman SJ, Taylor RC, Heniningfield JE. 1994. Nicotine and Smoking: A review of effects on human performance. *Experimental and Clinical Psychopharmacology*, 2: 345-395.
- Hoffmann D, Hecht SS. 1985. Nicotine-derived N-nitrosamines and tobacco related cancer: current status and future directions. *Cancer Research*, 45: 935-944.

23. Jendrassik L, Grof P. 1938. Colorimetric Method of Determination of bilirubin. *Biochemistry Z*, 297:81-82.
24. Julien RM. 1996. A primer of drug action. 7th edn. New York, W.H. Freeman & Co.
25. Meyer AW, Klesges RC, Winders SE, Ward KD, Peterson BA, Eck IH. 1997. Are weight concerns predictive of smoking cessation? A Prospective analysis. *Journal of consulting and clinical psychology*, 6: 448-452.
26. Mitra G, Poddar MK, Ghosh JJ. 1980. Effect of tobacco-leaf extract administration on liver, lung, intestine and serum enzymes. *Toxicology and Allied Pharmacology*, 52(2): 262-266.
27. Molowo SO. 2000. Toxicological Implications of ceftriaxone administration in rats. *Nigerian Journal of Biochemistry*, 15: 33-38.
28. Mustapha AA, Enemali MO, Olose M, Owuna G, Ogaji JO, Idris MM, Aboh VO. 2014. Phytoconstituents and Antibacterial efficacy of Mango (*Mangifera indica*) leave extracts. *Journal of Medicinal Plants Studies*, 2(5): 19-23.
29. Mustapha AA. 2014. Medicinal plants with possible anti- HIV activities: A Review. *International Journal of Medicinal Plants*, 106: 439-453.
30. Nweze CC, Ubhenin AE, Lay TU, Muhammed AU (2017) Hypoglycemic, hepatoprotective and hypolipidemic effect of pleurotus ostreatus in Alloxan-induced hyperglycemic rats. *Tropical Journal of Natural Product Research*, 1(4): 163-167.
31. Luka CD, Olatunde A, Tijjani H, Olisa-Enewe IA (2013) Effect of Aqueous extract of Phaseolus vulgaris L. (Red kidney beans) on alloxan induced Diabetic Wistar Rats. *Int. J. Sci. Inv Today*, 2(4): 292-30.
32. RecGCC. 1972. Colorimetric method for serum alkaline phosphatase determination. *Journal of Clinical Biochemistry*, 10(20): 182-184.
33. Reitman S, Frankel SA. 1957. Colorimetric method for the determination of serum oxaloacetic and glutamic pyruvate transaminase. *American Journal of clinical Pathology*, 28: 56-63.
34. Ren N, Timko MP. 2001. AFLP analysis of genetic polymorphism and evolutionary relationships among cultivated and wild Nicotiana species. *Genome*, 44(4): 559-571.
35. Russel MAH, Tarvis M, Feyerabend C. 1980. A view age for Snuff. *Lancet*, 1: 474-475.
36. Searcy RL and Berquist LM. (1960) A new colour reaction for the quantification of serum cholesterol. *clinchem Acta*, 5: 192-199.
37. Stepanov I, Villalta PW, A. Knezevich A, Jensen J, Hatsukami D, Hecht SS. 2010. Analysis of 23 polycyclic aromatic hydrocarbons in smokeless tobacco by gas chromatography-mass spectrometry. *Chemical Research in Toxicology*, 23(1): 66-73.
38. Tikian SM, Conover MB, Tikian AG. 1979. Clinical implications of Laboratory test. Pp. 3-44, 117-132, 154-159, London, St. Louis Toronto.
39. Trinder P. (1969) Mono reagent enzymatic glucose in clinical chemistry (W.B. Saunders, Philadelphia/LONDON) Pp. 24-27.
40. Ugbor CI, Okogun GRA, Okonkwo LO, Eze NC, Asogwa BE, Ebo JO, Maduagwu GN, Ekoh SN. 2013. The effect of tobacco snuff consumption on liver enzymes. *International Journal of Herbs and Pharmacological Research*, 2(2): 20-27.
41. Ureme SO, Ibeagha ID, Maduka IG, Ibeagbulam OG. 2007. The concentrations of methaemoglobin, carboxyhaemoglobin and some haematological parameters in tobacco snuff addicts in Igbo of Nigeria. *Nigerian Journal of Physiological Sciences*, 22 (1-2): 27-30.
42. Vik T, Jacobsen G, Vetten L, Bakkevig LS. 1996. Pre and post-natal growth in children of women who smoked in pregnancy. *Early Human Development*, 45: 245-255.
43. Whitby LG, Smith AF, Beckett GJ. 1989. Lecture notes on clinical chemistry. 4th Edn. pp. 38-178, Oxford London, Blackwell Scientific Publications.
44. Wolf PL. 1978. Clinical Significance of an increase or decrease serum alkaline phosphatase. *Archive of Pathology and Laboratory Medicine*, 102: 497-501.
45. Yakubu MT, Bilbis LS, Lawal M, Akanji MA (2003) Effect of Repeated Administration of Sildenafil Citrate on Selected Enzyme Activities of

Liver and Kidney of Male Albino Rats *Nigerian Journal of Pure Applied Sci.* 18: 1395-1400.

Article Keywords

Nicotianatabacum, Hepatotoxicity, hyperlipidemia, hyperglycemia

Funding:

This study has not received any external funding.

Conflict of Interest:

The authors declare that there are no conflicts of interests.

Data and materials availability:

All data associated with this study are present in the paper.

Authors Declaration

The Authors hereby declare that this study is original research work and that we shall be liable for any claims relating to the content of this article.

Article History

Received: 29 June 2018

Accepted: 06 August 2018

Published: August 2018

Citation

Nweze Chibuzo Carole, Nweze OtitoAdaeze. Hepatotoxicity, Hyperglycemic and hyperlipidemic effect of Aqueous Leaf Extract of *Nicotianatabacum* in male Wistar albino rats. *Drug Discovery*, 2018, 12, 54-58

Publication License



© The Author(s) 2018. Open Access. This article is licensed under a Creative Commons Attribution License 4.0 (CC BY 4.0).

General Note



Article is recommended to print as color digital version in recycled paper. *Save trees, save nature*